

09/512,515
DIALOG

Set	Items	Description
S1	51049	IMMUNOPEROXIDASE
S2	152076	NONSPECIFIC
S3	207	NORMAL (W) GOAT (W) SERUM
S4	51049	S1
S5	30	S2 (S) S3
S6	3	S1 AND S5
S7	75672	GOAT
S8	6087	GOAT (S) MOUSE
S9	380283	INDIRECT
S10	20	S1 AND S8 AND S9
S11	9	RD (unique items)
S12	67	S1 AND S8
S13	47	S12 NOT S10
S14	25	RD (unique items)
S15	24512	REHYDRAT?
S16	68174	FORMALIN
S17	935010	ALCOHOL
S18	169761	PARAFFIN?
S19	1	S15 AND S17 AND S18 AND S1
S20	11	S15 AND S18 AND S17
S21	10	RD (unique items)
S22	88424	IMMUNOSTAINING
S23	15	S22 AND S18 AND S15
S24	5	RD (unique items)

DIALOG

14/3,AB,K/7 (Item 7 from file: 155)

DIALOG(R)File 155:MEDLINE(R)

05813907 86197622 PMID: 3009605

Characterization of epithelial membrane antigen expression in human mammary epithelium by ultrastructural immunoperoxidase cytochemistry.

Petersen OW; van Deurs B

journal of histochemistry and cytochemistry (UNITED STATES) Jun 1986,

34 (6) p801-9, ISSN 0022-1554 Journal Code: IDZ

Languages: ENGLISH

Document type: Journal Article

Record type: Completed

Ultrastructural immunocytochemistry was used to analyze cell surface distribution and intracellular localization of milk fat globule membrane antigen (MFGM-A) in cryosections from human breast carcinomas and benign breast biopsy specimens. The specimens were fixed in formaldehyde and frozen. Cryostat sections were cut at 15 micron, incubated with mouse monoclonal antibody to MFGM-A, and then with a peroxidase-conjugated goat anti- mouse antibody. After glutaraldehyde fixation, the sections were incubated with diaminobenzidine-H2O2 and further processed for electron microscopy. MFGM-A was specific for epithelial cells. MFGM-A staining was strictly confined to the apical surface membrane of normal ductal epithelium, never involving basolateral membranes below the tight junctions. In normal epithelial cells, MFGM-A was readily detected in cisternae of the endoplasmic reticulum (ER), but only to a lesser extent in Golgi complexes and presumptive secretory vesicles. In carcinoma cells, surface staining for MFGM-A was either distributed in a non-polarized manner on the entire cell surface or else was totally absent. In some carcinoma cells without surface-associated MFGM-A, very pronounced intracellular MFGM-A staining was seen in the ER, in the nuclear envelope, and in annulate lamellae. The observations on MFGM-A expression were supported by studies on a cell culture model system.

DIALOG

6/3,AB,K/2 (Item 2 from file: 155)
DIALOG(R) File 155: MEDLINE(R)

03905999 83203481 PMID: 6342576

Use of the immunoperoxidase method on paraffin sections]

Primenenie immunoperoksidaznogo metoda na parafinovykh srezakh.

Babaev VR; Shchetnikova LA

Arkhiv patologii (USSR) 1983, 45 (1) p76-8, ISSN 0004-1955

Journal Code: 80E

Languages: RUSSIAN

Document type: Journal Article

Record type: Completed

Using an indirect **immunoperoxidase** technique, the methods of fixation and treatment of paraffin sections were explored for subsequent identification of myosin of smooth and skeletal muscle cells, actin, filamin, and immunoglobulins. Bouin's fluid and ethanol were found to be the most adequate fixators for immunomorphological analysis. Pronase treatment of paraffin sections increased the intensity of specific reactions considerably. Incubation of paraffin sections with **normal goat serum** and hydrogen peroxide decreased **nonspecific** staining of the sections.